

## CLAIM AMENDMENTS:

1. (currently amended) A solid state nuclear magnetic resonance (NMR) method for investigating a sample material that contains protons H and also spin-1/2 hetero nuclei X, the method comprising the steps of:
  - a) increasing an equilibrium polarization of X;
  - b) suppressing proton magnetization;
  - c) transferring polarization from X to H using a radio frequency (RF) pulse sequence which effects transfer between the nuclei X and spatially proximate protons H utilizing a dipole coupling constant  $D_{XH}$ , wherein polarization transfer depends only weakly on couplings of nuclei X to spatially distant protons and only weakly on couplings among the protons themselves;
  - d) recording proton signals under a line narrowing condition, wherein the sample material is rotated at a magic angle (MAS = magic angle spinning);
  - e) repeating steps a) through d) several times while varying an experimental parameter which is clearly physically associated with a polarization transfer process; and
  - f) determining a dipole coupling constant  $D_{XH}$  by analyzing variations in intensity of proton signals recorded in step d), wherein a ratio between a number of H nuclei to a number of X nuclei is larger than or equal to 10:1.
2. (cancelled)
3. (currently amended) The method of ~~claim 2~~ claim 1, wherein said ratio is larger than or equal to 100:1.

## 3

4. (original) The method of claim 1, wherein the X nuclei in the sample have natural abundance.
5. (original) The method of claim 1, wherein the X nuclei have a gyromagnetic ratio of  $\gamma(X) \leq \gamma(^{13}\text{C})$ .
6. (original) The method of claim 1, wherein the X nuclei comprise  $^{15}\text{N}$ .
7. (original) The method of claim 1, wherein the X nuclei comprise  $^{13}\text{C}$ .
8. (original) The method of claim 1, wherein the X nuclei comprise  $^{29}\text{Si}$ .
9. (original) The method of claim 1, wherein a polarization transfer from H to X is effected in step a).
10. (original) The method of claim 1, wherein a cross-polarization is applied in step a).
11. (original) The method of claim 1, wherein a field gradient pulse is applied in step b).
12. (original) The method of claim 1, wherein two radio frequency pulses are applied in step b) having a rotary resonance recoupling condition.
13. (original) The method of claim 1, wherein a chemical shift of the X nuclei is encoded between steps b) and c) under proton decoupling in a time interval  $t_1$ .
14. (original) The method of claim 1, wherein a TEDOR/REPT sequence is applied in step c), with a time interval  $t_1'$  being an experimental parameter which is clearly physically associated with the transfer

## 4

process, said time interval  $t_1'$  being used between a  $90^\circ$  pulse on X and a  $90^\circ$  pulse on H for encoding the dipole coupling constant  $D_{XH}$ .

15. (original) The method of claim 13, wherein a TEDOR/REPT sequence is applied in step c), with a time interval  $t_1'$  being an experimental parameter which is clearly physically associated with the transfer process, said time interval H being used between a  $90^\circ$  pulse on X and a  $90^\circ$  pulse on H for encoding the dipole coupling constant  $D_{XH}$ .
16. (original) The method of claim 15, wherein steps a) through d) are carried out several times in succession, wherein  $t_1$  and  $t_1'$  are simultaneously incremented.
17. (original) The method of claim 16, wherein  $t_1$  and  $t_1'$  are incremented with different time increments.
18. (original) The method of claim 1, wherein a TEODOR/REPT sequence is applied in step c), wherein a time interval  $t_1'$  between a  $90^\circ$  pulse on X and a  $90^\circ$  pulse on H is fixed and a number of rotor-synchronized  $180^\circ$  pulses is varied as an experimental parameter which is clearly physically associated with a polarization transfer process, wherein intensities in resulting spectra for different numbers of rotor-synchronized  $180^\circ$  pulses are used to determine dipole coupling constants  $D_{XH}$ .
19. (original) The method of claim 1, wherein a TEDOR/REPT sequence is applied in step c), and a time interval  $t_1'$  between a  $90^\circ$  pulse on X and a  $90^\circ$  pulse on H is fixed and a time difference between rotor-synchronized  $180^\circ$  pulses on X relative to rotor-synchronized  $180^\circ$  pulses on H is varied as an experimental parameter which is clearly physically associated with a polarization transfer process, wherein

## 5

dipole coupling constants  $D_{XH}$  are determined from spectra extracted for different time differences.

20. (original) The method of claim 19, wherein a chemical shift of the X nuclei is encoded between steps b) and c) under proton decoupling in a time interval  $t_1$  and steps a) through d) are carried out several times in succession, wherein both  $t_1$  and a time difference between the rotor-synchronized  $180^\circ$  pulses on X and rotor-synchronized  $180^\circ$  pulses on H are simultaneously incremented.
21. (original) The method of claim 20, wherein  $t_1$  and said time difference are incremented with different time increments.
22. (original) The method of claim 1, wherein transfer in step c) is effected by a Lee-Goldburg cross-polarization whose time duration is varied as an experimental parameter which is clearly physically associated with a polarization transfer process.
23. (original) The method of claim 1, wherein rapid rotation at the magic angle with a rotation frequency which is larger than or equal to 25 kHz (fast MAS) is only effected in step d).
24. (original) The method of claim 1, wherein rotation at the magic angle is supported by radio frequency pulses in step d).
25. (original) The method of claim 1, wherein rotation at the magic angle is supported by pulsed spin locking in step d).
26. (original) The method of claim 1, wherein the method determines X-H binding separations.

## 6

27. (original) The method of claim 26, wherein said binding separations are of hydrogen bridges.
28. (original) The method of claim 1, wherein the method is applied to determine a structure of a peptide chain.